

Prevention and Treatment of Alzheimer Disease and Aging: Antioxidants

Quan Liu^{1,3,*}, Fang Xie², Raj Rolston¹, Paula I. Moreira^{1,4}, Akihiko Nunomura⁵, Xiongwei Zhu¹, Mark A. Smith¹ and George Perry^{1,6,*}

¹Departments of Pathology and ²Pharmacology, Case Western Reserve University, Cleveland Ohio 44106 USA; ³Department of Ophthalmology, University of California, San Diego, San Diego, California 92093 USA; ⁴Center for Neuroscience and Cell Biology of Coimbra, University of Coimbra, 3004-517 Coimbra, Portugal; ⁵Department of Psychiatry and Neurology, Asahikawa Medical College, Asahikawa 078-8510, Japan; ⁶College of Sciences, University of Texas at San Antonio, San Antonio, Texas 78249 USA

Abstract: There is considerable evidence showing that oxidative damage is one of the earliest neuronal and pathological changes of Alzheimer disease and many, if not all, of the etiological and pathological causes of the disease are related, directly or indirectly, to free radical production and oxidative damage. Here we summarize the current body of knowledge suggestive that oxidative damage is, if not the key factor, certainly a major factor in Alzheimer disease. As such, therapeutic modalities encompassing antioxidants may be an effective approach to the treatment of neurodegenerative diseases and delay the aging process.

Key Words: Antioxidant, calorie restriction, estrogen, free radical, glutathione, oxidation, oxidative stress, sirtuin, therapy, vitamin C, vitamin E.

GENERAL INFORMATION ABOUT ALZHEIMER DISEASE

Alzheimer disease (AD) was first reported by Dr. Alois Alzheimer, a German doctor, in 1907 at a neurology conference, where he described a 51-year-old woman with rapid memory degeneration who died with severe dementia several years later [1,2]. Although the disease was once considered rare, it is now established as the leading cause of dementia. According to the American Health Assistance Foundation and the Alzheimer's Association, there are an estimated 4 million diseased individuals in the United States and 18 million worldwide, with as many as 350,000 individuals being diagnosed with the disease each year. In the US alone, annual expenses exceed US \$70-\$100 billion [3]. The disease is still not curable, with current clinical therapy of cholinesterase inhibitors/NMDA receptor antagonists such as Reminyl (galantamine), Aricept (donepezil hydrochloride), Exelon (rivastigmine) and Namenda (memantine), which offer little more than short-term palliative effects.

AD involves the parts of the brain that control thought, memory, and language. After two decades of intensive study, during which much more information has been obtained, the cause of AD is still a mystery. Currently, there are several major theories of AD, such as amyloid- β (A β) toxicity [4], tauopathy [5], inflammation [6,7], oxidative stress [8-13], all of which have been vigorously argued in the literature.

CLINICAL FACTS OF ALZHEIMER DISEASE

The risk of AD varies from 12% to 19% for women over the age of 65 years and 6% to 10% for men [14] and rises

exponentially with age, such that up to 47% of individuals over age of 80 develop AD [15]. On average, AD patients live about 8 years after initial diagnosis, although the disease can last for as long as 20 years. The areas of the brain that control memory and thinking skills are affected first but, as the disease progresses, neurons in other regions of the brain are also affected. Eventually, the patient with AD will need complete care. The physical and emotional burden of the disease is borne by the patient and family members until the patient's death.

PATHOLOGY OF ALZHEIMER DISEASE

Two distinctive hallmark lesions found in the brains of patients with AD are senile plaques and neurofibrillary tangles (NFTs) (reviewed in [2]) which were identified by the use of silver-staining techniques [16]. In addition, other neuropathological changes associated with the disease include neuronal and dendritic loss, neuropil threads, dystrophic neurites, granulovacuolar degeneration, Hirano bodies, cerebrovascular amyloid, and atrophy of the brain [2].

Senile plaques are spherical extracellular lesions, 10-200 μ m in diameter, with a central core made of bundles of 6-10 nm A β [2]. In the peripheral region of the senile plaques, A β and amyloid- β protein precursor (A β PP), tau, and neurofilament proteins are reported [17].

NFTs, the major intracellular protein aggregation found in AD brains, are located primarily in the cerebral cortex, especially in the large pyramidal neurons in the hippocampal and frontotemporal regions [18]. NFTs are composed of bundles of paired helical filaments (PHF), the major component of which is the microtubule-associated protein tau [19, 20]. Moreover, neurofilament proteins are also reported in NFTs [17,21]. In PHF, tau is abnormally hyperphosphorylated [5,22,23], ubiquitinated [24-26], oxidized [8,27-29], truncated [30] and aggregated into filaments [5,31,32]. The hyperphosphorylation of tau is thought to render it unable to

*Address correspondence to these authors at the Department of Ophthalmology, University of California, San Diego, 9500 Gilman Drive # 0946, La Jolla, California 92093-0946 USA; Tel: 858-534-8824; Fax: 858-534-1625; E-mail: quliu@ucsd.edu

College of Sciences, University of Texas at San Antonio, 6900 North Loop 1604 West, San Antonio, Texas 78249 USA; Tel: 210-458-4450; Fax: 210-458-4445; E-mail: george.perry@utsa.edu

bind to microtubules and therefore unable to promote or maintain microtubule assembly [33], although *in vivo*, while microtubules are disrupted, this has no relation to NFT [34]. The resistance to proteolytic degradation of hyperphosphorylated tau may play a key role in neurofibrillary degeneration in AD patients [35,36].

While the pathological hallmarks are the basis for current diagnostic standards, whether they represent the initial causes or the consequences of disease is hotly contested [37-39].

ETIOLOGY OF ALZHEIMER DISEASE

Only about 5% of all AD cases have an early onset and are related to genetic mutations of *presenilin 1*, *presenilin 2* or the *AβPP* genes [40]. Indeed, the majority, approximately 95%, of all AD patients are sporadic, late-onset cases where the major risk factors are aging and *apolipoprotein E4* (*ApoE4*) polymorphisms [41,42]. In both familial and sporadic cases of disease, there is accumulating evidence indicating a major role for free radicals and oxidative stress in disease pathogenesis and pathophysiology [11,12].

Age

Age is the single greatest risk factor for AD and the disease rarely occurs in people under 60 years. Thereafter, AD affects 10-15% of individuals over 65 years old and up to 47% of individuals over the age of 80 [15]. This predominance of age as a major cause in AD etiology indicates that age-related events are closely involved in the development of the disease. While the processes of aging that are involved in AD pathogenesis are not fully understood, two likely candidates are altered cholinergic function and oxidative stress. The former, decreases of cholinergic neurons with age and disease [43] is the basis for therapy with three currently used drugs that stabilize acetylcholine levels in neurons. The latter, oxidative stress is discussed in detail below.

Oxidative Stress

As one of the leading proposed causes of aging [44], free radical damage and oxidative stress are also thought to play a major role in the pathogenesis of AD. Oxidative stress is a potential source of damage to DNA, lipids, sugars and proteins within cells. Any imbalance between the intracellular production of free radicals/reactive oxygen species (ROS) and antioxidant defense mechanisms results in oxidative stress [8,11,12]. Since neurons have an age-related decrease in the capacity to compensate for redox imbalance, even minor cellular stresses have the ability to lead to irreversible injury and, as such, contribute to the pathogenesis of neurodegenerative diseases. ROS, including free radicals, are the primary mediators of oxidative injury and cause damage to lipids, sugars, DNA/RNA and amino acid side-chains [13, 45]. Markers for oxidative damage (carbonyls, HNE, MDA and more) may increase in neurodegenerative diseases and aging, but whether they can be used as quantifiable markers for disease condition is unclear.

Genetics

The greatest correlated genetic factor for the development of AD are polymorphisms of the *ApoE* gene, such that 50% of patients with AD patients have at least one *ApoE4* allele [41,42]. Further, although familial AD only accounts

for a small percentage (~5%) of total AD cases, many have argued that mutations in *AβPP* and *presenilin 1* and *2* are critical genetic factors in the AD pathogenesis and in Aβ production [46,47].

Apolipoprotein E Gene Polymorphism: Polymorphisms of the *ApoE* gene are found to correlate with onset and risk of developing AD. *ApoE* is an abundant 34-kDa glycoprotein that is synthesized and secreted mainly by astrocytes and microglia in the central nervous system (CNS). It is well established that *ApoE*, and especially the *E4* allele of *ApoE*, is a major genetic risk factor for the more common, late-onset form of AD [48,49]. The influence of *ApoE* genotype on AD seems to operate *via* multiple mechanisms. For example, polymorphisms are a determinant of brain Aβ burden in individuals affected with AD [50,51]. Additionally, apolipoproteins have been suggested to act as antioxidants, with the *ApoE4* allele being less effective in this role [52] so that increased oxidative damage is found in specific brain regions of AD patients with the *ApoE E4* genotype [53].

Amyloid-β Protein Precursor

Aβ protein is the major component of senile plaque cores and is derived from the precursor protein, AβPP. AβPP is encoded on chromosome 21 (21q11-22) [54,55]. The normal function of AβPP is unknown, but it is involved in several broad physiological functions in neurons. Mutations in AβPP appear to change AβPP processing and while initially this was thought to lead to increases in Aβ, thus increasing the extracellular protein aggregation [56,57], more recent reports actually show decreases in Aβ [58]. Transgenic mice that overexpress mutant *AβPP* show overproduction of Aβ protein, senile plaque formation and synaptic deficits without NFTs pathology, indicating a key pathological role for mutant AβPP protein [59,60]. The current data finds that AβPP mutation only accounts for a very small percentage of AD cases, 0.1-0.15% of total AD cases.

Presenilins 1 and 2

The majority (~70%) of early-onset familial AD cases are associated with mutations in two genes, *presenilin 1* and *presenilin 2*, located on chromosomes 14 and 1, respectively [61]. Over 50 different pathogenic mutations in *presenilin 1* gene and 3 mutations in *presenilin 2* gene have been described [62]. There is considerable homology between the gene products of *presenilin 1* and *presenilin 2*, which are transmembrane proteins of 463 and 448 amino acids respectively, with six and nine hydrophobic membrane-spanning domains [61]. The physiological functions of these two proteins are unknown but may be involved in the Notch receptor pathway [63]. Other possible roles include ion channel, protein processing, or cellular trafficking functions [64]. In AD, it is thought mutations in these proteins are associated with AD by affecting the processing of AβPP [65].

Tauopathy

Hyperphosphorylation of tau makes it more resistant to proteolytic degradation, which may play a key role in neurofibrillary degeneration in AD patients [35,36]. Tau aggregation was, until quite recently, viewed as being deleterious. However, more recent evidence indicates it is a consequence of neurodegeneration. In fact, tau aggregation may be an

adaptive change for the neurons to absorb oxidative stress [29,38,66,67]. Consistent with this notion, tau phosphorylation and aggregation and NFT epitopes have been shown experimentally to be both consequences of oxidative stress and post-translational oxidation of tau [29,31,68-71].

Other Factors

Vascular risk factors: Hyperlipidemia, hypertension, diabetes, and related factors of heart disease or stroke have been identified as putative antecedents to AD [72].

Down Syndrome

Adults with Down syndrome develop the neuropathological changes of AD by age 40, but not all patients become demented. The risk of AD in families with a history of Down syndrome is increased 2-3 fold.

Alcohol

Individuals who drink red wine in moderate amounts daily are less likely to develop AD than either heavier drinkers or abstainers [73]. The risk reduction associated with alcohol is possibly related to its anti-inflammatory and antioxidant properties or its effects on lipid metabolism by components such as resveratrol [74].

Education and Early Life Experience

Several studies show that the risk of AD among poorly educated individuals or individuals in poor living condition is significantly higher than that among well-educated persons [75,76].

Smoking

Smokers have a 2-4 fold increase in risk of AD, particularly those individuals without an ApoE4 allele [77,78].

Head Injury

There is an increase of the risk of AD related with traumatic head injury [79].

Anti-Inflammatory Drugs

AD was found to be less frequent among individuals who used anti-inflammatory agents [80].

Hormone Replacement

The use of estrogen by postmenopausal women has been associated with a decreased risk of AD [81,82]. Women using hormone replacement had about a 50% reduction in disease risk with benefit only to those taking estrogen in the peri-menopausal period. While the exact mechanism for this is unclear, recent evidence points to the feedback effect of estrogen on luteinizing hormone [83-92].

OXIDATIVE STRESS IS THE KEY FACTOR IN ALZHEIMER DISEASE

Evidence Supporting Oxidative Damage in Alzheimer Disease

The histopathological and the experimental evidence, which support the impact of oxidative damage in the pathogenesis of AD, are outlined below [8,10,12,93].

- 1) Oxidative damage is an earlier change compared to other pathological manifestations of the disease [94,95].
- 2) Increased levels of oxidative damage are found in post-mortem tissue of AD, including oxidative modifications of lipid, protein, DNA and sugar [27,96-99].
- 3) Increased response to oxidative stress or compensatory defense exaggeration can be seen in the affected brain regions [100-103].
- 4) High amounts of metal ions can be seen in AD brain [96,104,105].
- 5) Altered mitochondria functions are commonly observed [106,107].
- 6) Formation of a specific age-associated oxidative crosslink of proteins [108].

Free Radical Theory of Aging and Free Radical Production

Aging is the inevitable decline in physiologic functions that occurs over time and, for all living organisms, ends in death. At least four major theories of aging have been proposed to explain most or all of the physiological and pathological changes of biological aging:

- 1) The free radical theory of aging;
- 2) The mitochondrial theory of aging;
- 3) The cross-link theory of aging;
- 4) The membrane hypothesis of aging;

Professor Denham Harman, the founder of the free radical theory of aging, has defined aging as the increased probability of death as the age of an organism increases, and diverse adverse physiologic changes accumulate [44]. Free radicals, the highly reactive small oxygen-containing molecules, undeniably play major roles in not only the free radical theory of aging but also in the mitochondrial theory, membrane theory and cross-link theory as well or is a common feature among all of them.

A molecule carrying an unpaired electron, which makes it extremely reactive and ready to acquire an electron in any way possible, is termed a free radical. In the process of acquiring an electron, the free radical will attach itself to another molecule, thereby modifying it biochemically [109]. However, as free radicals acquire an electron from the other molecules, they either convert these molecules into other free radicals, or break down or alter their chemical structure. Thus, free radicals are capable of damaging virtually any biomolecule, including proteins, sugars, fatty acids and nucleic acids [110]. Free radical damage to long-lived biomolecules such as collagen, elastin, DNA, polysaccharides, lipids that make up the membranes of cells and organelles, blood vessel walls and lipofuscins is thought of as a major contributor to cell death [111].

The most common free radicals include superoxide, hydroxyl, hydroperoxyl, alkoxy, peroxy and nitric oxide radical. Other non-free radical molecules, such as singlet oxygen, hydrogen peroxide (H₂O₂), and hypochlorous acid

(HOCl), are similar but not real free radicals. Together, the free radicals and free radical mimics are called ROS.

Free radicals have extremely short half-lives ranging from nanosecond to seconds. The shortest is only one nanosecond (10^{-9} sec) for hydroxyl radical and the longest half-life is 1-10 seconds for nitric oxide radical [112]. The half-life dictates the intrinsic properties of the damaging effects of the free radicals, whether they can travel far enough to reach other cellular compartments or just attack the most nearby molecules. The further they can travel, the broader the range of molecules and organelles they can damage.

A wide range of major diseases closely related to free radical damage, such as cancer, heart/artery disease, essential hypertension, AD, cataracts, diabetes, Parkinson's disease, arthritis and inflammatory disease, as well as aging itself, are now believed to be caused in part or entirely by free radical damage [111,113].

Sources of Free Radicals

There are more than six primary sources of free radicals formed endogenously within living organisms.

The major source of free radicals and oxidants is through the respiratory generation of ATP using oxygen. [113-115]; the second source of free radical production is the peroxisomal oxidation of fatty acids, which generates H_2O_2 as a by-product [114,115]; the third source is cytochrome P450 enzymes [115]; the fourth, and preventable, source of free radical production is from chronic inflammatory cells which use a mixture of oxidants to overcome infection by phagocytosis [110,114,115]; the fifth source is from other enzymes capable of generating oxidants under normal or pathological conditions [116]; the sixth source is various biomolecules including thiols, hydroquinones, flavins, catecholamines, pterins and hemoglobin, may spontaneously auto-oxidize and produce superoxide radicals [110]. Recent research indicates that A β could induce peptide fragmentation and free radical production [56,117].

Many exogenous sources, such as environmental radiation (sunlight), polluted urban air, cigarette smoke, iron and copper salts, some phenolic compounds found in many plant foods, and various drugs [110,114] could contribute to free radical production.

Antioxidant Systems

Endogenous Antioxidants

To protect against free radical-induced cellular damage, cells have endogenous defense mechanisms to quench free radicals that include enzymatic antioxidant systems and cellular molecules.

SOD, catalase, and glutathione peroxidase are three primary enzymes involved in direct elimination of active oxygen species (superoxide radical and H_2O_2). In addition, glutathione reductase, glucose-6-phosphate dehydrogenase, and cytosolic GST are secondary enzymes. The latter function to decrease peroxide levels or to maintain a steady supply of metabolic intermediates like glutathione (GSH) and NADPH for optimum functioning of primary antioxidant enzymes [118,119].

Many cellular molecules are active antioxidants in the body. For example, GSH, ascorbate (vitamin C), α -tocopherol (vitamin E), β -carotene, NADPH, uric acid, bilirubin, selenium, mannitol, benzoate, the iron-binding protein transferrin, dihydrolipoic acid, melatonin, plasma protein thiol, and reduced CoQ10 are all involved in protecting the body from ROS and their byproducts produced during normal cellular metabolism. Of these, GSH is the most significant component that directly quenches ROS such as lipid peroxides (like hydroxynonenal) and plays major role in xenobiotic metabolism. Exposure to high levels of xenobiotics causes GSH to be exhausted in the process of xenobiotic neutralization and it is therefore less available to serve as an antioxidant. GSH is also important in maintaining ascorbate (vitamin C) and α -tocopherol (vitamin E) in their reduced form so they may function as antioxidants to quench free radicals [120-122].

Exogenous Antioxidants from the Diet

The most widely studied dietary antioxidants are vitamin C, vitamin E, and β -carotene. Vitamin C is considered the most important water-soluble antioxidant in extracellular fluids, as it is capable of neutralizing ROS in the aqueous phase before lipid peroxidation is initiated. Vitamin E is a major lipid-soluble antioxidant, and is the most effective chain-breaking antioxidant within the cell membrane, where it protects membrane fatty acids from lipid peroxidation. β -carotene and other carotenoids also provide antioxidant protection to lipid rich tissues. Fruits and vegetables are major sources of vitamin C and carotenoids. Whole grains, cereals, and high quality vegetable oils, are major sources of vitamin E [123-125].

The Vulnerability of The Nervous System

The nervous system – including the brain, spinal cord, and peripheral nerves – is rich in both unsaturated fatty acids and iron. The double bonds in unsaturated fatty acids make them a vulnerable target for free radicals, and this, coupled with the high aerobic metabolic activity in neurons, makes the nervous system particularly susceptible to oxidative damage. The high level of iron, while it may be essential, particularly during brain development, facilitates oxidative stress *via* iron-catalyzed formation of ROS [126,127]. In addition, those brain regions that are rich in the catecholamines, adrenaline, noradrenaline and dopamine, are exceptionally vulnerable to free radical generation. Catecholamines can induce free radicals through either spontaneous breakdown (auto-oxidation) or by being metabolized by endogenous enzymes such as monoamine oxidase. One such region of the brain is the substantia nigra, where a connection has been established between antioxidant depletion (including GSH) and tissue degeneration [11].

There is an increase in markers of oxidative stress in major neurodegenerative diseases [8,27,28,96-98,101,128,129] and substantial evidence that oxidative stress is a cause, or at least the initial change, in the pathogenesis of AD [94].

ANTIOXIDANT CLINICAL TRIALS AND STUDIES FOR THE TREATMENT OF ALZHEIMER DISEASE

Current Clinical Drugs in Use

Therapy with acetylcholinesterase inhibitors, including drugs such as Reminyl, Aricept and Exelon, which aim to

stabilize acetylcholine levels in the synaptic cleft to maintain neurotransmission, is based on the hypothesis that cholinergic dysfunction in the process of aging contributes to the development of AD. The newly-developed drug, Memantine, an NMDA receptor antagonist, blocks glutamate-mediated excitotoxicity. All drugs currently in clinical usage are reviewed elsewhere [130-132].

Antioxidant Therapy Development

The stages of free radical production may be arbitrarily divided into 1) conditions prior to their formation, 2) free radical formation and 3) adduction. The different types of antioxidant therapy are based on their intervention at different points in the stages of free radical formation. Summarized below are current strategies for developing antioxidant therapy for AD.

Compounds or Methods That Prevent/Reduce Formation of Free Radicals

Modulation of SOD, Peroxidase and Catalase or Using Their Mimics

Overexpression of human Cu-Zn SOD in transgenic mice showed reduced oxidative damage in brain [133] and improved cognitive functions in aged rodents [134]. Overexpression of glutathione peroxidase in transgenic mice also showed antioxidative function and rescued homocysteine-induced endothelial dysfunction [135]. Moreover, the mimics of SOD and catalase have cytoprotective effects in AD

model systems [136] and prolong lifespan in *C. elegans* [137]. It is reported that SOD, peroxidase and catalase activity is reduced with age and in some pathological conditions [138]. Therapy aimed at compensating for loss of activity of these enzymes is a promising approach to AD therapy.

Iron Chelators

A β PP transgenic mice treated with a Cu/Zn chelator showed improvement in general health parameters and a reduction of brain A β deposition [139]. Because copper and zinc play a major role in A β toxicity and nerve cell death *via* ROS generation, chelator therapy is, in effect, antioxidant [140-142]. In one study, 48 presumed AD patients treated with desferrioxamine, a transition metal chelator, (250 mg per day), showed this class of compounds to be effective in preventing AD progression [143]. Recently, desferrioxamine and others, as FDA-preapproved drugs, were shown to limit A β protein secretion in cell culture [144]. More iron chelators are under investigation and show beneficial effect in AD treatment [145].

Caloric Restriction

Studies of caloric restriction in rodents show an attenuation of age-related deficits in learning and memory [146] and dramatically extends the life-span and reduces the incidence of age-related disease in rodents and monkeys [147,148]. The mechanism of the beneficial effect of caloric restriction is not clearly understood but is most likely *via* overall reduc-

Table 1. Classification of Antioxidant Therapy to Different Stages of Free Radical Production

Classifications	Importance	References
<u>Compounds or Methods That Prevent/Reduce Formation of Free Radicals</u>		
Modulation of SOD, peroxidase and catalase or using their mimics	*	[133-138]
Iron chelators	****	[139-143]
Caloric restriction	***	[146-153]
<u>Compounds That Directly Scavenge Free Radicals</u>		
Tocopherols (vitamin E), ascorbate (vitamin C)	*****	[121,156-164,199]
Estrogen	****	[157,165-173]
Glutathione	***	[174-178]
Others:		
Serotonin (5-hydroxytryptamine), quercetin, idebenone	**	[180-183,185]
TMP	**	[167]
Uric acid	*	[155]
Aromatic amino and imino compounds	*	[189,190]
Carotene, flavonoids and other polyphenols, retinol and other polyenes	*	[154]
<u>Compounds That Can Limit the Extent of Damage to Detoxify or Prevent the Formation of ROS Adducts</u>		
Tenilsetam	*	[191,192]
Reparative enzymes (methione sulfoxide reductase)	*	[193-198]

* - frequency or relative numbers of studies.

tion in levels of oxidative stress, including in the brain [149]. In humans, a low daily calorie intake is associated with a reduced risk for AD [150]. In addition, the incidence of AD is lower in countries with low per capita food consumption compared to countries with high per capita food consumption [151,152]. Reducing caloric intake as a preventive measure in populations at high risk for AD could be combined with other AD treatment. The development of chemical mimics for caloric restriction, such as resveratrol and sirtuins [153,154], may make caloric restriction for normal people more easily attainable in the future.

Compounds That Directly Scavenge Free Radicals

Various compounds have the ability to quench free radicals by reacting with them directly. These compounds include tocopherols (vitamin E) and other monophenols, ascorbate (vitamin C), carotene, flavonoids and other polyphenols, GSH ratio (GSH/GSSH), retinol and other polyenes, arylamines and indoles, ebselen and other selenium-containing compounds, mimics of catalase/SOD, etc. The major antioxidants in the group of direct antioxidants are the chain-breaking antioxidants, such as phenols and carotenes.

Vitamin E and Vitamin C

Vitamin E has been found in rats, to prevent neuronal cell death induced by hypoxia followed by oxygen reperfusion and to prevent neuronal damage from reactive nitrogen species [155]. Both vitamin E and β -carotene, by reducing oxidative stress, protect rat neurons from exposure to ethanol [156]. In an experimental model of diabetes-caused neurovascular dysfunction β -carotene, followed by vitamins E and C, was found to protect cells effectively [157]. Vitamin E can rescue the neuronal cytotoxicity induced by aluminum in A β PP transgenic mice and reduce A β deposition in the brain by reducing isoprostane levels [95].

A significant amount of research with dietary vitamins E and C has been done in humans. In a multicenter, double blind, placebo-controlled study on 341 patients with moderately severe AD, a daily dose of approximately 1350 mg (2000 IU) vitamin E led to a slight delay of AD progression, providing the first evidence for vitamin E as prophylaxis and treatment for AD [158]. In keeping with these results, other studies with vitamins C and E in AD patients have shown that antioxidants might have a protective effect against AD [159,160].

In one later study, 815 non-demented individuals were evaluated based on their intake of the antioxidants vitamins C and E and β -carotene. Results showed that there is a significant difference in the incidence of AD between those taking vitamin E and those who are not [161]. In another large-scale study involving 5396 non-demented individuals, it was reported that a high intake of vitamins C and E significantly reduces the risk of AD [162]. Similarly, in a 5-year follow-up with 1367 non-demented individuals over 65 years of age in France, Commenges and colleagues [163] found that an intake of flavonoids significantly reduced the risk of dementia.

While promising, many studies have also reported negative results to disagree with the effectiveness of vitamin E and vitamin C intake as detailed in other reviews [164].

Estrogen

The general neuroprotective effects of estrogen (17 β -estradiol) have been the subject of much research. Generally, estrogen through interaction with its receptors ER α and ER β , acts as a trophic factor in the nervous system by altering gene transcription [165]. At the same time, in a manner more closely related to direct antioxidants, estrogen can have antioxidant activity independent of estrogen receptors. The structure of estrogen is much like β -tocopherol in that both molecules contain a phenolic radical scavenging moiety and a lipophilic carbohydrate moiety. In general, phenolic A ring estrogens have been shown to be powerful inhibitors of lipid peroxidation in various cell-free test-tube experiments [166-168]. By changing the phenolic character of estrogen 17 β -estradiol, its antioxidant activities are lost, supporting the theory that estrogens are direct antioxidants because of their phenolic ring structure [169,170]. Furthermore, this antioxidant activity of estrogens and other phenols is strictly related to the structural prerequisites and not dependent on the interaction of these compounds with cellular estrogen receptors [167]. Meanwhile it has been shown that estrogens are strong antioxidants in different oxidative stress-induced cell degeneration models [169-172].

One thing to be considered is the optimum concentration of estrogen needed to achieve the antioxidant effect. Normally, estrogen is present in nanomolar concentrations *in vivo* and, in most *in vitro* studies, estrogen's antioxidant effect is achieved at a significantly higher concentration range of 1-10 μ moles. Therefore, it remains to be determined how the antioxidant effect of estrogen can be achieved therapeutically with a safe pharmacologic dose.

It was recently shown that HRT, in the form of estrogen plus progestin, administered as a therapeutic agent in a WHI clinical trial was shown to increase the risk for probable dementia in postmenopausal women aged 65 years or older [173]. The investigators responsible for this study hypothesize that the negative effect of estrogen and progestin may be linked to the increased risk of stroke that was also reported in the estrogen/progestin treatment group, as the relationship between microinfarcts in the brain and susceptibility to AD is likely related, yet currently not well characterized. While this may indeed partially explain the results of the WHI clinical trial, it is only when the role of the other hormones of the hypothalamic-pituitary-gonadal axis during the climacteric years and beyond is taken into account that the results of the WHI clinical trial can be fully and accurately explained. For instance, it is crucial when interpreting the results of this study to recognize that the hormones of the hypothalamic-pituitary-gonadal axis have been in disequilibrium for decades in all of the women who participated in the WHI clinical trial, so if a lack of estrogen does indeed play a role in AD pathogenesis, these women have been exposed to this disease-promoting hormonal environment for years if not decades by the time the estrogen/progestin treatment was administered. This is evidenced by the fact that reports of probable dementia appeared within the first year of the study in both the treatment and placebo groups. Therefore, it is likely predictable that the administration of estrogen/progestin in these aged women was not only unable to restore the proper functioning of the hypothalamic-pituitary-gonadal

axis, but that the influx of exogenous hormones actually served to exacerbate the disease process.

Glutathione

GSH, the most abundant intracellular non-protein thiol, is the main factor which directly quenches free radicals *in vivo*. It has been shown that the level of GSH is decreased in cortical areas and in the hippocampus of patients with AD as compared with controls [174-176]. The level of GSH in red blood cells decreases with aging and in patients with AD [177]. In the healthy cell, oxidized glutathione (GSSG) rarely exceeds 10% of total cellular GSH. Therefore, the ratio of GSH/GSSG can be used as a useful indicator for oxidative status *in vivo* [178]. GSH depletion may be the ultimate factor determining vulnerability to oxidant attack. N-acetyl-cysteine (NAC), a precursor of GSH which has already been approved by the U.S. Food and Drug Administration for treatment of acetaminophen toxicity, may be an effective strategy to increase GSH and spare brain degeneration in AD patients, although this remains to be tested.

Other Direct Antioxidants

Compounds such as serotonin (5-hydroxytryptamine), flavanoids, quercetin, and simple alkylphenols have been shown to prevent membrane lipid peroxidation and protect neuronal cells against oxidative cell death *in vitro* [179,180].

2,4,6-trimethylphenol (TMP) is also a potent antioxidant [167]. Additionally, being a small compound, TMP would readily cross the blood-brain barrier, thus meeting the most critical requirement for drugs used in the treatment of neurodegenerative diseases. The protective potential of this compound is currently being tested experimentally in various animal models of acute neurodegeneration.

In two large clinical studies, administration of idebenone, a compound structurally similar to ubiquinone, has been reported to significantly reduce disease progression in a dose-dependent fashion [181,182]. Some *in vivo* studies in animals as well as *in vitro* studies have demonstrated a protective effect of idebenone in neuronal death [183]. More recent studies argued the effectiveness of idebenone in AD treatment [184,185], but another study showed that idebenone is better than tacrine in benefit-risk ratio in AD treatment [186]. Whether idebenone acts by modulating mitochondrial metabolic function or directly as a radical scavenger is still an open question.

Uric acid, an endogenous antioxidant, was also found to prevent ischemia-induced oxidative neuronal damage in rats [155]. In addition, cannabidiol is more effective than either vitamin C or E in protecting against glutamate neurotoxicity [187]. It has been demonstrated that the antioxidant activity of cannabinoid compounds is, similar to estrogens, exclusively dependent on the presence of a phenolic group and is independent of the cannabinoid receptor [188].

Different aromatic amino and imino compounds (e.g., phenothiazine, phenoxazine, iminostilbene) are another group of direct antioxidants. Aromatic amines and imines are effective against oxidative glutamate toxicity, GSH depletion, and H₂O₂ toxicity in different cell culture systems [189]. With

half-maximal effective concentrations ranging from 20-75 nM, these compounds were experimentally proven to be more effective than common standard phenolic antioxidants. These results provide a structural basis and rationale for the development of new antioxidant drugs [189,190].

Compounds That Can Limit the Extent of Damage to, Detoxify or Prevent the Formation of ROS Adducts

There are yet another group of compounds that can detoxify the formed ROS adducts and repair the damage they produce. These include NAC, GSH, 2-oxo-thiazolidine-4-carboxylate, carnitine, creatine, lipoic acid (thioctic acid), ubiquinone and idebenone, etc.

The compound tenilsetam, a cognition-enhancing drug, is sometimes used to treat AD patients. Its mechanism of action is unclear but, based on *in vitro* and *in vivo* evidence, it is believed to inhibit protein glycation [191,192]. Since generation of advanced glycation endproducts is a major manifestation of oxidative stress in AD [8,98,129] glycation inhibition is thought to be neuroprotective.

Many amino acid residues of proteins are susceptible to oxidation by various forms of ROS. Oxidatively-modified proteins accumulate during aging and in a number of age-related diseases. There is an increase in oxidation of the S-containing amino acids methionine and cysteine in AD patients [193]. However, unlike oxidation of other amino acid residues, the oxidation of these two amino acids can be repaired by corresponding enzymes, methionine sulfoxide reductases (MSR), thioredoxin reductase (TrxR), thioredoxin (Trx), and NADPH. The level of MSR is decreased with aging and in AD and other neurodegenerative diseases [193]. Also, mutation in the MSR gene in yeast, bacteria, and mice as well as its overexpression in yeast, neuronal PC-12 cells, human T cells and *Drosophila* has been correlated with increased antioxidant capacity and the prolonged life span of these organisms [194-198].

CONCLUSIONS AND FUTURE PERSPECTIVES

It has been well established that oxidative damage of cellular molecules plays an important role in neurodegenerative disorders. Furthermore, it is clear that oxidative damage is not simply a byproduct or end product of neuronal degenerative processes but, more likely, the direct initiation factor in neurodegeneration.

Currently, even with the huge amount of data produced and increase in knowledge, there is much skepticism regarding the likelihood of success with antioxidant therapy in AD. The only promising results so far are from the trial of vitamin E therapy in moderately severe AD [158,199]. It would be difficult for most of the presently known compounds with antioxidant activity to pass through the blood brain barrier. There is much scope for research to identify smaller antioxidant molecules that would more readily pass through the blood brain barrier and/or non-toxic or inert compounds that would carry antioxidant drugs from the bloodstream into the brain. Additionally, it is imperative that future trials use combinations, rather than single antioxidants to facilitate redox cycling as well as maximize bioavailability to different cellular compartments.

REFERENCES

- [1] Alzheimer, A. *Allg. Zeitschr. Psychiatr.*, **1907**, *64*, 146.
- [2] Alzheimer, A.; Stelzmann, R.A.; Schnitzlein, H.N.; Murtagh, F.R. *Clin. Anat.*, **1995**, *8*, 429.
- [3] Mayeux, R. *Annu. Rev. Neurosci.*, **2003**, *26*, 81.
- [4] Selkoe, D.J. *Neurol. Clin.*, **2000**, *18*, 903.
- [5] Lee, V.M.; Goedert, M.; Trojanowski, J.Q. *Annu. Rev. Neurosci.*, **2001**, *24*, 1121.
- [6] McGeer, P.L.; McGeer, E.G. *Neurobiol. Aging*, **2001**, *22*, 799.
- [7] Weiner, H.L.; Selkoe, D.J. *Nature*, **2002**, *420*, 879.
- [8] Smith, M.A.; Sayre, L.M.; Monnier, V.M.; Perry, G. *Trends Neurosci.*, **1995**, *18*, 172.
- [9] Markesbery, W.R. *Free Radic. Biol. Med.*, **1997**, *23*, 134.
- [10] Christen, Y. *Am. J. Clin. Nutr.*, **2000**, *71*, 621S.
- [11] Perry, G.; Nunomura, A.; Hirai, K.; Zhu, X.; Perez, M.; Avila, J.; Castellani, R.J.; Atwood, C.S.; Aliev, G.; Sayre, L.M.; Takeda, A.; Smith, M.A. *Free Radic. Biol. Med.*, **2002**, *33*, 1475.
- [12] Perry, G.; Castellani, R.J.; Hirai, K.; Smith, M.A. *J. Alzheimers Dis.*, **1998**, *1*, 45.
- [13] Picklo, M.J.; Montine, T.J.; Amarnath, V.; Neely, M.D. *Toxicol. Appl. Pharmacol.*, **2002**, *184*, 187.
- [14] Seshadri, S.; Wolf, P.A.; Beiser, A.; Au, R.; McNulty, K.; White, R.; D'Agostino, R.B. *Neurology*, **1997**, *49*, 1498.
- [15] Evans, D.A.; Funkenstein, H.H.; Albert, M.S.; Scherr, P.A.; Cook, N.R.; Chown, M.J.; Hebert, L.E.; Hennekens, C.H.; Taylor, J.O. *JAMA*, **1989**, *262*, 2551.
- [16] Alzheimer, A. *Zentralblatt. fur. Nervenkrankheiten*, **1906**, *25*, 1134.
- [17] Perry, G.; Lipphardt, S.; Mulvihill, P.; Kancherla, M.; Mijares, M.; Gambetti, P.; Sharma, S.; Maggiora, L.; Cornette, J.; Lobl, T.; Greenberg, B. *Lancet*, **1988**, *2*, 746.
- [18] Smith, M.A. *Int. Rev. Neurobiol.*, **1998**, *42*, 1.
- [19] Grundke-Iqbal, I.; Iqbal, K. *Prog. Clin. Biol. Res.*, **1989**, *317*, 745.
- [20] Lee, V.M.; Balin, B.J.; Otvos, L., Jr.; Trojanowski, J.Q. *Science*, **1991**, *251*, 675.
- [21] Liu, Q.; Raina, A.K.; Smith, M.A.; Sayre, L.M.; Perry, G. *Mol. Aspects Med.*, **2003**, *24*, 305.
- [22] Grundke-Iqbal, I.; Iqbal, K.; Tung, Y.C.; Quinlan, M.; Wisniewski, H.M.; Binder, L.I. *Proc. Natl. Acad. Sci. USA*, **1986**, *83*, 4913.
- [23] Trojanowski, J.Q.; Lee, V.M. *FASEB J.*, **1995**, *9*, 1570.
- [24] Mori, H.; Kondo, J.; Ihara, Y. *Science*, **1987**, *235*, 1641.
- [25] Perry, G.; Friedman, R.; Shaw, G.; Chau, V. *Proc. Natl. Acad. Sci. USA*, **1987**, *84*, 3033.
- [26] Iqbal, K.; Grundke-Iqbal, I. *Int. Psychogeriatr.*, **1997**, *9* (Suppl. 1), 289.
- [27] Sayre, L.M.; Zelasko, D.A.; Harris, P.L.; Perry, G.; Salomon, R.G.; Smith, M.A. *J. Neurochem.*, **1997**, *68*, 2092.
- [28] Takeda, A.; Smith, M.A.; Avila, J.; Nunomura, A.; Siedlak, S.L.; Zhu, X.; Perry, G.; Sayre, L.M. *J. Neurochem.*, **2000**, *75*, 1234.
- [29] Liu, Q.; Smith, M.A.; Avila, J.; DeBernardis, J.; Kansal, M.; Takeda, A.; Zhu, X.; Nunomura, A.; Honda, K.; Moreira, P.I.; Oliveira, C.R.; Santos, M.S.; Shimohama, S.; Aliev, G.; de la Torre, J.; Ghanbari, H.A.; Siedlak, S.L.; Harris, P.L.; Sayre, L.M.; Perry, G. *Free Radic. Biol. Med.*, **2005**, *38*, 746.
- [30] Gamblin, T.C.; Chen, F.; Zambrano, A.; Abraha, A.; Lagalwar, S.; Guillozet, A.L.; Lu, M.; Fu, Y.; Garcia-Sierra, F.; LaPointe, N.; Miller, R.; Berry, R.W.; Binder, L.I.; Cryns, V.L. *Proc. Natl. Acad. Sci. USA*, **2003**, *100*, 10032.
- [31] Perez, M.; Hernandez, F.; Gomez-Ramos, A.; Smith, M.; Perry, G.; Avila, J. *Eur. J. Biochem.*, **2002**, *269*, 1484.
- [32] Lovestone, S.; McLoughlin, D.M. *J. Neurol. Neurosurg. Psychiatry*, **2002**, *72*, 152.
- [33] Baudier, J.; Cole, D.R. In *Advances in Behavioral Biology, Vol. 34, Alterations in the Neuronal Cytoskeleton in Alzheimer Disease*; Perry, G., Ed.; Plenum Press: New York, **1987**; pp. 25.
- [34] Cash, A.D.; Aliev, G.; Siedlak, S.L.; Nunomura, A.; Fujioka, H.; Zhu, X.; Raina, A.K.; Vinters, H.V.; Tabaton, M.; Johnson, A.B.; Paula-Barbosa, M.; Avila, J.; Jones, P.K.; Castellani, R.J.; Smith, M.A.; Perry, G. *Am. J. Pathol.*, **2003**, *162*, 1623.
- [35] Eidenmuller, J.; Fath, T.; Hellwig, A.; Reed, J.; Sontag, E.; Brandt, R. *Biochemistry*, **2000**, *39*, 13166.
- [36] Cras, P.; Smith, M.A.; Richey, P.L.; Siedlak, S.L.; Mulvihill, P.; Perry, G. *Acta Neuropathol. (Berl.)*, **1995**, *89*, 291.
- [37] Smith, M.A.; Casadesus, G.; Joseph, J.A.; Perry, G. *Free Radic. Biol. Med.*, **2002**, *33*, 1194.
- [38] Lee, H.G.; Perry, G.; Moreira, P.I.; Garrett, M.R.; Liu, Q.; Zhu, X.; Takeda, A.; Nunomura, A.; Smith, M.A. *Trends Mol. Med.*, **2005**, *11*, 164.
- [39] Lee, H.G.; Zhu, X.; Nunomura, A.; Perry, G.; Smith, M.A. *Curr. Alzheimer Res.*, **2006**, *3*, 75.
- [40] Selkoe, D.J. *Physiol. Rev.*, **2001**, *81*, 741.
- [41] Ashford, J.W. *J. Mol. Neurosci.*, **2004**, *23*, 157.
- [42] Teter, B. *J. Mol. Neurosci.*, **2004**, *23*, 167.
- [43] Whitehouse, P.J.; Price, D.L.; Clark, A.W.; Coyle, J.T.; DeLong, M.R. *Ann. Neurol.*, **1981**, *10*, 122.
- [44] Harman, D. *J. Gerontol.*, **1956**, *11*, 298.
- [45] Honda, K.; Smith, M.A.; Zhu, X.; Baus, D.; Merrick, W.C.; Tartakoff, A.M.; Hattier, T.; Harris, P.L.; Siedlak, S.L.; Fujioka, H.; Liu, Q.; Moreira, P.I.; Miller, F.P.; Nunomura, A.; Shimohama, S.; Perry, G. *J. Biol. Chem.*, **2005**, *280*, 20978.
- [46] Hardy, J.A.; Higgins, G.A. *Science*, **1992**, *256*, 184.
- [47] Hardy, J.; Selkoe, D.J. *Science*, **2002**, *297*, 353.
- [48] Saunders, A.M.; Schmechel, K.; Breitner, J.C.; Benson, M.D.; Brown, W.T.; Goldfarb, L.; Goldgaber, D.; Manwaring, M.G.; Szymanski, M.H.; McCown, N.; Dole, K.C.; Schmechel, D.E.; Strittmatter, W.J.; Pericak-Vance, M.A.; Roses, A.D. *Lancet*, **1993**, *342*, 710.
- [49] Rebeck, G.W.; Reiter, J.S.; Strickland, D.K.; Hyman, B.T. *Neuron*, **1993**, *11*, 575.
- [50] Schmechel, D.E.; Saunders, A.M.; Strittmatter, W.J.; Crain, B.J.; Hulette, C.M.; Joo, S.H.; Pericak-Vance, M.A.; Goldgaber, D.; Roses, A.D. *Proc. Natl. Acad. Sci. USA*, **1993**, *90*, 9649.
- [51] Strittmatter, W.J.; Weisgraber, K.H.; Huang, D.Y.; Dong, L.M.; Salvesen, G.S.; Pericak-Vance, M.; Schmechel, D.; Saunders, A.M.; Goldgaber, D.; Roses, A.D. *Proc. Natl. Acad. Sci. USA*, **1993**, *90*, 8098.
- [52] Lauderback, C.M.; Kanski, J.; Hackett, J.M.; Maeda, N.; Kindy, M.S.; Butterfield, D.A. *Brain Res.*, **2002**, *924*, 90.
- [53] Ramassamy, C.; Averill, D.; Beffert, U.; Theroux, L.; Lussier-Cacan, S.; Cohn, J.S.; Christen, Y.; Schoofs, A.; Davignon, J.; Poitrier, J. *Neurobiol. Dis.*, **2000**, *7*, 23.
- [54] Tanzi, R.E.; Gusella, J.F.; Watkins, P.C.; Bruns, G.A.; St George-Hyslop, P.; Van Keuren, M.L.; Patterson, D.; Pagan, S.; Kurnit, D.M.; Neve, R.L. *Science*, **1987**, *235*, 880.
- [55] St George-Hyslop, P.H.; Tanzi, R.E.; Polinsky, R.J.; Haines, J.L.; Nee, L.; Watkins, P.C.; Myers, R.H.; Feldman, R.G.; Pollen, D.; Drachman, D.; Growdon, J.; Bruni, A.; Foncin, J.-F.; Salmon, D.; Frommelt, P.; Amaducci, L.; Sorbi, S.; Piacentini, S.; Stewart, G.D.; Hobbs, W.J.; Conneally, P.M.; Gusella, J.F. *Science*, **1987**, *235*, 885.
- [56] Selkoe, D.J. *J. Alzheimers Dis.*, **2001**, *3*, 75.
- [57] Selkoe, D.J. *Science*, **1997**, *275*, 630.
- [58] Bentahir, M.; Nyabi, O.; Verhamme, J.; Tolia, A.; Horre, K.; Wiltfang, J.; Esselmann, H.; De Strooper, B. *J. Neurochem.*, **2006**, *96*, 732.
- [59] Hsiao, K.; Chapman, P.; Nilsen, S.; Eckman, C.; Harigaya, Y.; Younkin, S.; Yang, F.; Cole, G. *Science*, **1996**, *274*, 99.
- [60] Games, D.; Adams, D.; Alessandrini, R.; Barbour, R.; Berthelette, P.; Blackwell, C.; Carr, T.; Clemens, J.; Donaldson, T.; Gillespie, F.; Guido, T.; Hagopian, S.; Johnson-Wood, K.; Khan, K.; Lee, M.; Leibowitz, P.; Lieberburg, I.; Little, S.; Masliah, E.; McConlogue, L.; Montoya-Zavala, M.; Mucke, L.; Paganini, L.; Penniman, E.; Power, M.; Schenk, D.; Seubert, P.; Snyder, B.; Soriano, F.; Tan, H.; Vitale, J.; Wadsworth, S.; Wolozin, B.; Zhao, J. *Nature*, **1995**, *373*, 523.
- [61] Rogaev, E.I.; Sherrington, R.; Rogaeva, E.A.; Levesque, G.; Ikeda, M.; Liang, Y.; Chi, H.; Lin, C.; Holman, K.; Tsuda, T.; Mar, L.; Sorbi, S.; Nacmias, B.; Piacentini, S.; Amaducci, L.; Chumakov, I.; Cohen, D.; Lannfelt, L.; Fraser, P.E.; Rommens, J.M.; St. George-Hyslop, P.H. *Nature*, **1995**, *376*, 775.
- [62] Checler, F. *C. R. Acad. Sci. III*, **1999**, *322*, 1033.
- [63] Sherrington, R.; Rogaev, E.I.; Liang, Y.; Rogaeva, E.A.; Levesque, G.; Ikeda, M.; Chi, H.; Lin, C.; Li, G.; Holman, K.; Tsuda, T.; Mar, L.; Foncin, J.-F.; Bruni, A.C.; Montesti, M.P.; Sorbi, S.; Rainero, I.; Pinessi, L.; Nee, L.; Chumakov, I.; Pollen, D.; Brookes, A.; Sanseau, P.; Polinsky, R.J.; Wasco, W.; Da Silva, H.A.R.; Haines, J.L.; Pericak-Vance, M.A.; Tanzi, R.E.; Roses, A.D.; Fraser, P.E.; Rommens, J.M.; St. George-Hyslop, P.H. *Nature*, **1995**, *375*, 754.

- [64] Czech, C.; Tremp, G.; Pradier, L. *Prog. Neurobiol.*, **2000**, *60*, 363.
- [65] Scheuner, D.; Eckman, C.; Jensen, M.; Song, X.; Citron, M.; Suzuki, N.; Bird, T.D.; Hardy, J.; Hutton, M.; Kukull, W.; Larson, E.; Levy-Lahad, E.; Viitanen, M.; Peskind, E.; Poorkaj, P.; Schellenberg, G.; Tanzi, R.; Wasco, W.; Lannfelt, L.; Selkoe, D.; Younkin, S. *Nat. Med.*, **1996**, *2*, 864.
- [66] Liu, Q.; Lee, H.G.; Honda, K.; Siedlak, S.L.; Harris, P.L.; Cash, A.D.; Zhu, X.; Avila, J.; Nunomura, A.; Takeda, A.; Smith, M.A.; Perry, G. *Biochim. Biophys. Acta*, **2005**, *1739*, 211.
- [67] Takeda, A.; Perry, G.; Abraham, N.G.; Dwyer, B.E.; Kutty, R.K.; Laitinen, J.T.; Petersen, R.B.; Smith, M.A. *J. Biol. Chem.*, **2000**, *275*, 5395.
- [68] Perez, M.; Cuadros, R.; Smith, M.A.; Perry, G.; Avila, J. *FEBS Lett.*, **2000**, *486*, 270.
- [69] Zhu, X.; Rottkamp, C.A.; Boux, H.; Takeda, A.; Perry, G.; Smith, M.A. *J. Neuropathol. Exp. Neurol.*, **2000**, *59*, 880.
- [70] Zhu, X.; Castellani, R.J.; Takeda, A.; Nunomura, A.; Atwood, C.S.; Perry, G.; Smith, M.A. *Mech. Ageing Dev.*, **2001**, *123*, 39.
- [71] Zhu, X.; Raina, A.K.; Rottkamp, C.A.; Aliev, G.; Perry, G.; Boux, H.; Smith, M.A. *J. Neurochem.*, **2001**, *76*, 435.
- [72] Breteler, M.M. *Neurobiol. Aging*, **2000**, *21*, 153.
- [73] Orgogozo, J.M.; Dartigues, J.F.; Lafont, S.; Letenneur, L.; Comenges, D.; Salamon, R.; Renaud, S.; Breteler, M.B. *Rev. Neurol. (Paris)*, **1997**, *153*, 185.
- [74] de la Lastra, C.A.; Villegas, I. *Mol. Nutr. Food Res.*, **2005**, *49*, 405.
- [75] Stern, Y.; Gurland, B.; Tatemichi, T.K.; Tang, M.X.; Wilder, D.; Mayeux, R. *JAMA*, **1994**, *271*, 1004.
- [76] Hall, K.S.; Gao, S.; Unverzagt, F.W.; Hendrie, H.C. *Neurology*, **2000**, *54*, 95.
- [77] Merchant, C.; Tang, M.X.; Albert, S.; Manly, J.; Stern, Y.; Mayeux, R. *Neurology*, **1999**, *52*, 1408.
- [78] Ott, A.; Slioter, A.J.; Hofman, A.; van Harskamp, F.; Witteman, J.C.; Van Broeckhoven, C.; van Duijn, C.M.; Breteler, M.M. *Lancet*, **1998**, *351*, 1840.
- [79] Plassman, B.L.; Havlik, R.J.; Steffens, D.C.; Helms, M.J.; Newman, T.N.; Drosdick, D.; Phillips, C.; Gau, B.A.; Welsh-Bohmer, K.A.; Burke, J.R.; Guralnik, J.M.; Breitner, J.C. *Neurology*, **2000**, *55*, 1158.
- [80] Weggen, S.; Eriksen, J.L.; Das, P.; Sagi, S.A.; Wang, R.; Pietrzik, C.U.; Findlay, K.A.; Smith, T.E.; Murphy, M.P.; Bulter, T.; Kang, D.E.; Marquez-Sterling, N.; Golde, T.E.; Koo, E.H. *Nature*, **2001**, *414*, 212.
- [81] Baldereschi, M.; Di Carlo, A.; Lepore, V.; Bracco, L.; Maggi, S.; Grigoletto, F.; Scarlato, G.; Amaducci, L. *Neurology*, **1998**, *50*, 996.
- [82] Waring, S.C.; Rocca, W.A.; Petersen, R.C.; O'Brien, P.C.; Tangalos, E.G.; Kokmen, E. *Neurology*, **1999**, *52*, 965.
- [83] Casadesus, G.; Smith, M.A.; Zhu, X.; Aliev, G.; Cash, A.D.; Honda, K.; Petersen, R.B.; Perry, G. *J. Alzheimers Dis.*, **2004**, *6*, 165.
- [84] Casadesus, G.; Zhu, X.; Atwood, C.S.; Webber, K.M.; Perry, G.; Bowen, R.L.; Smith, M.A. *Curr. Drug Targets CNS Neurol. Disord.*, **2004**, *3*, 281.
- [85] Casadesus, G.; Atwood, C.S.; Zhu, X.; Hartzler, A.W.; Webber, K.M.; Perry, G.; Bowen, R.L.; Smith, M.A. *Cell Mol. Life Sci.*, **2005**, *62*, 293.
- [86] Casadesus, G.; Shukitt-Hale, B.; Stellwagen, H.M.; Zhu, X.; Lee, H.G.; Smith, M.A.; Joseph, J.A. *Nutr. Neurosci.*, **2004**, *7*, 309.
- [87] Casadesus, G.; Webber, K.M.; Atwood, C.S.; Pappolla, M.A.; Perry, G.; Bowen, R.L.; Smith, M.A. *Biochim. Biophys. Acta*, **2006**, *1762*, 447.
- [88] Casadesus, G.; Milliken, E.L.; Webber, K.M.; Bowen, R.L.; Lei, Z.; Rao, C.V.; Atwood, C.S.; Perry, G.; Keri, R.A.; Smith, M.A. *Mol. Cell. Endocrinol.*, **2006**, in press.
- [89] Webber, K.M.; Bowen, R.; Casadesus, G.; Perry, G.; Atwood, C.S.; Smith, M.A. *Acta Neurobiol. Exp. (Wars)*, **2004**, *64*, 113.
- [90] Webber, K.M.; Casadesus, G.; Perry, G.; Atwood, C.S.; Bowen, R.; Smith, M.A. *Alzheimer Dis. Assoc. Disord.*, **2005**, *19*, 95.
- [91] Webber, K.M.; Casadesus, G.; Marlatt, M.W.; Perry, G.; Hamlin, C.R.; Atwood, C.S.; Bowen, R.L.; Smith, M.A. *Ann. N Y Acad. Sci.*, **2005**, *1052*, 201.
- [92] Webber, K.M.; Casadesus, G.; Zhu, X.; Obrenovich, M.E.; Atwood, C.S.; Perry, G.; Bowen, R.L.; Smith, M.A. *Curr. Pharm. Des.*, **2006**, *12*, 691.
- [93] Markesbery, W.R. *Arch. Neurol.*, **1999**, *56*, 1449.
- [94] Nunomura, A.; Perry, G.; Aliev, G.; Hirai, K.; Takeda, A.; Balraj, E.K.; Jones, P.K.; Ghanbari, H.; Wataya, T.; Shimohama, S.; Chiba, S.; Atwood, C.S.; Petersen, R.B.; Smith, M.A. *J. Neuropathol. Exp. Neurol.*, **2001**, *60*, 759.
- [95] Pratico, D.; Clark, C.M.; Liun, F.; Rokach, J.; Lee, V.Y.; Trojanowski, J.Q. *Arch. Neurol.*, **2002**, *59*, 972.
- [96] Smith, M.A.; Richey Harris, P.L.; Sayre, L.M.; Beckman, J.S.; Perry, G. *J. Neurosci.*, **1997**, *17*, 2653.
- [97] Smith, M.A.; Rudnicka-Nawrot, M.; Richey, P.L.; Praprotnik, D.; Mulvihill, P.; Miller, C.A.; Sayre, L.M.; Perry, G. *J. Neurochem.*, **1995**, *64*, 2660.
- [98] Smith, M.A.; Taneda, S.; Richey, P.L.; Miyata, S.; Yan, S.D.; Stern, D.; Sayre, L.M.; Monnier, V.M.; Perry, G. *Proc. Natl. Acad. Sci. USA*, **1994**, *91*, 5710.
- [99] Smith, M.A.; Perry, G.; Richey, P.L.; Sayre, L.M.; Anderson, V.E.; Beal, M.F.; Kowall, N. *Nature*, **1996**, *382*, 120.
- [100] Pappolla, M.A.; Omar, R.A.; Kim, K.S.; Robakis, N.K. *Am. J. Pathol.*, **1992**, *140*, 621.
- [101] Smith, M.A.; Kutty, R.K.; Richey, P.L.; Yan, S.D.; Stern, D.; Chader, G.J.; Wiggert, B.; Petersen, R.B.; Perry, G. *Am. J. Pathol.*, **1994**, *145*, 42.
- [102] Premkumar, D.R.; Smith, M.A.; Richey, P.L.; Petersen, R.B.; Castellani, R.; Kutty, R.K.; Wiggert, B.; Perry, G.; Kalara, R.N. *J. Neurochem.*, **1995**, *65*, 1399.
- [103] Smith, M.A.; Richey, P.L.; Kutty, R.K.; Wiggert, B.; Perry, G. *Mol. Chem. Neuropathol.*, **1995**, *24*, 227.
- [104] Lovell, M.A.; Robertson, J.D.; Teesdale, W.J.; Campbell, J.L.; Markesbery, W.R. *J. Neurol. Sci.*, **1998**, *158*, 47.
- [105] Sayre, L.M.; Perry, G.; Harris, P.L.; Liu, Y.; Schubert, K.A.; Smith, M.A. *J. Neurochem.*, **2000**, *74*, 270.
- [106] Swerdlow, R.H.; Khan, S.M. *Med. Hypotheses.*, **2004**, *63*, 8.
- [107] Hirai, K.; Aliev, G.; Nunomura, A.; Fujioka, H.; Russell, R.L.; Atwood, C.S.; Johnson, A.B.; Kress, Y.; Vinters, H.V.; Tabaton, M.; Shimohama, S.; Cash, A.D.; Siedlak, S.L.; Harris, P.L.; Jones, P.K.; Petersen, R.B.; Perry, G.; Smith, M.A. *J. Neurosci.*, **2001**, *21*, 3017.
- [108] Smith, M.A.; Sayre, L.M.; Anderson, V.E.; Harris, P.L.; Beal, M.F.; Kowall, N.; Perry, G. *J. Histochem. Cytochem.*, **1998**, *46*, 731.
- [109] Allen, P.C. *Poult. Sci.*, **1997**, *76*, 814.
- [110] Leibovitz, B.E.; Siegel, B.V. *J. Gerontol.*, **1980**, *35*, 45.
- [111] Harman, L.S.; Mottley, C.; Mason, R.P. *J. Biol. Chem.*, **1984**, *259*, 5606.
- [112] Priyadarsini, K.I.; Khopde, S.M.; Kumar, S.S.; Mohan, H. *J. Agric. Food Chem.*, **2002**, *50*, 2200.
- [113] Pierrefiche, G.; Laborit, H. *Exp. Gerontol.*, **1995**, *30*, 213.
- [114] Ames, B.N.; Shigenaga, M.K.; Hagen, T.M. *Proc. Natl. Acad. Sci. USA*, **1993**, *90*, 7915.
- [115] Beckman, K.B.; Ames, B.N. *Physiol. Rev.*, **1998**, *78*, 547.
- [116] Beal, M.F. *Ann. N Y Acad. Sci.*, **2003**, *991*, 120.
- [117] Hensley, K.; Carney, J.M.; Mattson, M.P.; Aksenova, M.; Harris, M.; Wu, J.F.; Floyd, R.A.; Butterfield, D.A. *Proc. Natl. Acad. Sci. USA*, **1994**, *91*, 3270.
- [118] Vendemiale, G.; Grattagliano, I.; Altomare, E. *Int. J. Clin. Lab. Res.*, **1999**, *29*, 49.
- [119] Singh, M.; Saini, H.K. *J. Cardiovasc. Pharmacol. Ther.*, **2003**, *8*, 135.
- [120] Meister, A. *Cancer Res.*, **1994**, *54*, 1969s.
- [121] Sies, H.; Stahl, W. *Am. J. Clin. Nutr.*, **1995**, *62*, 1315S.
- [122] Anderson, R.; Ramafi, G.; Theron, A.J. *Biochem. Pharmacol.*, **1996**, *52*, 341.
- [123] Jacob, R.A. *Nutr. Res.*, **1995**, *15*, 755-766.
- [124] Halliwell, B. *Lancet*, **1994**, *344*, 721.
- [125] Block, G. *Am. J. Clin. Nutr.*, **1991**, *53*, 270S.
- [126] Bauer, C.; Walcher, F.; Holanda, M.; Mertzluft, F.; Larsen, R.; Marzi, I. *J. Trauma*, **1999**, *46*, 886.
- [127] Andron, N.; Kaufman, J.B.; Clem, T.R.; Fass, R.; Shiloach, J. *Ann. N Y Acad. Sci.*, **1990**, *589*, 363.
- [128] Nunomura, A.; Perry, G.; Pappolla, M.A.; Wade, R.; Hirai, K.; Chiba, S.; Smith, M.A. *J. Neurosci.*, **1999**, *19*, 1959.
- [129] Castellani, R.J.; Harris, P.L.; Sayre, L.M.; Fujii, J.; Taniguchi, N.; Vitek, M.P.; Founds, H.; Atwood, C.S.; Perry, G.; Smith, M.A. *Free. Radic. Biol. Med.*, **2001**, *31*, 175.

- [130] Marlatt, M.W.; Webber, K.M.; Moreira, P.I.; Lee, H.G.; Casadesus, G.; Honda, K.; Zhu, X.; Perry, G.; Smith, M.A. *Curr. Med. Chem.*, **2005**, *12*, 1137.
- [131] Pietrzik, C.; Behl, C. *Int. J. Exp. Pathol.*, **2005**, *86*, 173.
- [132] Doody, R.S. *Geriatrics*, **2005**, *Suppl.*, 14.
- [133] Cardozo-Pelaez, F.; Song, S.; Parthasarathy, A.; Epstein, C.J.; Sanchez-Ramos, J. *Neurobiol. Aging*, **1998**, *19*, 311.
- [134] Succi, D.J.; Crandall, B.M.; Arendash, G.W. *Brain Res.*, **1995**, *693*, 88.
- [135] Weiss, N.; Zhang, Y.Y.; Heydrick, S.; Bierl, C.; Loscalzo, J. *Proc. Natl. Acad. Sci. USA*, **2001**, *98*, 12503.
- [136] Bruce, A.J.; Boling, W.; Kindy, M.S.; Peschon, J.; Kraemer, P.J.; Carpenter, M.K.; Holtsberg, F.W.; Mattson, M.P. *Nat. Med.*, **1996**, *2*, 788.
- [137] Melov, S.; Ravenscroft, J.; Malik, S.; Gill, M.S.; Walker, D.W.; Clayton, P.E.; Wallace, D.C.; Malfroy, B.; Doctrow, S.R.; Lithgow, G.J. *Science*, **2000**, *289*, 1567.
- [138] Andersen, H.R.; Jeune, B.; Nybo, H.; Nielsen, J.B.; Andersen-Ranberg, K.; Grandjean, P. *Age Ageing*, **1998**, *27*, 643.
- [139] Chemy, R.A.; Atwood, C.S.; Xilinas, M.E.; Gray, D.N.; Jones, W.D.; McLean, C.A.; Bamham, K.J.; Volitakis, I.; Fraser, F.W.; Kim, Y.; Huang, X.; Goldstein, L.E.; Moir, R.D.; Lim, J.T.; Beyreuther, K.; Zheng, H.; Tanzi, R.E.; Masters, C.L.; Bush, A.I. *Neuron*, **2001**, *30*, 665.
- [140] Smith, M.A.; Harris, P.L.; Sayre, L.M.; Perry, G. *Proc. Natl. Acad. Sci. USA*, **1997**, *94*, 9866.
- [141] Huang, X.; Cuijungeo, M.P.; Atwood, C.S.; Hartshorn, M.A.; Tyndall, J.D.; Hanson, G.R.; Stokes, K.C.; Leopold, M.; Multhaup, G.; Goldstein, L.E.; Scarpa, R.C.; Saunders, A.J.; Lim, J.; Moir, R.D.; Glabe, C.; Bowden, E.F.; Masters, C.L.; Fairlie, D.P.; Tanzi, R.E.; Bush, A.I. *J. Biol. Chem.*, **1999**, *274*, 37111.
- [142] Sayre, L.M.; Smith, M.A.; Perry, G. *Curr. Med. Chem.*, **2001**, *8*, 721.
- [143] Crapper McLachlan, D.R.; Dalton, A.J.; Kruck, T.P.; Bell, M.Y.; Smith, W.L.; Kalow, W.; Andrews, D.F. *Lancet*, **1991**, *337*, 1304.
- [144] Morse, L.J.; Payton, S.M.; Cuny, G.D.; Rogers, J.T. *J. Mol. Neurosci.*, **2004**, *24*, 129.
- [145] Liu, G.; Garrett, M.R.; Men, P.; Zhu, X.; Perry, G.; Smith, M.A. *Biochim. Biophys. Acta*, **2005**, *1741*, 246.
- [146] Ingram, D.K.; Weindruch, R.; Spangler, E.L.; Freeman, J.R.; Walford, R.L. *J. Gerontol.*, **1987**, *42*, 78.
- [147] Sohal, R.S.; Weindruch, R. *Science*, **1996**, *273*, 59.
- [148] Weindruch, R.; Lane, M.A.; Ingram, D.K.; Ershler, W.B.; Roth, G.S. *Aging (Milano)*, **1997**, *9*, 304.
- [149] Dubey, A.; Forster, M.J.; Lal, H.; Sohal, R.S. *Arch. Biochem. Biophys.*, **1996**, *333*, 189.
- [150] Mayeux, R.; Sano, M. *N. Engl. J. Med.*, **1999**, *341*, 1670.
- [151] Smith, M.A.; Petot, G.J.; Perry, G. *J. Alzheimers Dis.*, **1999**, *1*, 203.
- [152] Grant, W.B. *J. Alzheimers Dis.*, **1999**, *1*, 197.
- [153] Guarente, L.; Picard, F. *Cell*, **2005**, *120*, 473.
- [154] Delmas, D.; Jannin, B.; Latruffe, N. *Mol. Nutr. Food Res.*, **2005**, *49*, 377.
- [155] Yu, Z.F.; Bruce-Keller, A.J.; Goodman, Y.; Mattson, M.P. *J. Neurosci. Res.*, **1998**, *53*, 613.
- [156] Copp, R.P.; Wisniewski, T.; Hentati, F.; Larnaout, A.; Ben Hamida, M.; Kayden, H.J. *Brain Res.*, **1999**, *822*, 80.
- [157] Mitchell, J.H.; Collins, A.R. *Eur. J. Nutr.*, **1999**, *38*, 143.
- [158] Sano, M.; Ernesto, C.; Thomas, R.G.; Klauber, M.R.; Schafer, K.; Grundman, M.; Woodbury, P.; Growdon, J.; Cotman, C.W.; Pfeiffer, E.; Schneider, L.S.; Thal, L.J. *N. Engl. J. Med.*, **1997**, *336*, 1216.
- [159] Pitchumoni, S.S.; Doraiswamy, P.M. *J. Am. Geriatr. Soc.*, **1998**, *46*, 1566.
- [160] Flynn, B.L.; Ranno, A.E. *Ann. Pharmacother.*, **1999**, *33*, 188.
- [161] Morris, M.C.; Evans, D.A.; Bienias, J.L.; Tangney, C.C.; Bennett, D.A.; Aggarwal, N.; Wilson, R.S.; Scherr, P.A. *JAMA*, **2002**, *287*, 3230.
- [162] Engelhart, M.J.; Geerlings, M.I.; Ruitenberg, A.; van Swieten, J.C.; Hofman, A.; Wittteman, J.C.; Breteler, M.M. *JAMA*, **2002**, *287*, 3223.
- [163] Commenges, D.; Scotet, V.; Renaud, S.; Jacqmin-Gadda, H.; Barberger-Gateau, P.; Dartigues, J.F. *Eur. J. Epidemiol.*, **2000**, *16*, 357.
- [164] Boothby, L.A.; Doering, P.L. *Ann. Pharmacother.*, **2005**, *39*, 2073.
- [165] Koehler, K.F.; Helguero, L.A.; Haldosen, L.A.; Warner, M.; Gustafsson, J.A. *Endocr. Rev.*, **2005**, *26*, 465.
- [166] Sugioka, K.; Shimosegawa, Y.; Nakano, M. *FEBS Lett.*, **1987**, *210*, 37.
- [167] Moosmann, B.; Behl, C. *Proc. Natl. Acad. Sci. USA*, **1999**, *96*, 8867.
- [168] Behl, C. *MMW. Fortschr. Med.*, **2001**, *143* (Suppl. 2), 33.
- [169] Behl, C.; Skutella, T.; Lezoualc'h, F.; Post, A.; Widmann, M.; Newton, C.J.; Holsboer, F. *Mol. Pharmacol.*, **1997**, *51*, 535.
- [170] Behl, C.; Widmann, M.; Trapp, T.; Holsboer, F. *Biochem. Biophys. Res. Commun.*, **1995**, *216*, 473.
- [171] Dubal, D.B.; Kashon, M.L.; Pettigrew, L.C.; Ren, J.M.; Finklestein, S.P.; Rau, S.W.; Wise, P.M. *J. Cereb. Blood Flow Metab.*, **1998**, *18*, 1253.
- [172] Goodman, Y.; Bruce, A.J.; Cheng, B.; Mattson, M.P. *J. Neurochem.*, **1996**, *66*, 1836.
- [173] Shumaker, S.A.; Legault, C.; Rapp, S.R.; Thal, L.; Wallace, R.B.; Ockene, J.K.; Hendrix, S.L.; Jones, B.N., 3rd; Assaf, A.R.; Jackson, R.D.; Kotchen, J.M.; Wassertheil-Smoller, S.; Wactawski-Wende, J. *JAMA*, **2003**, *289*, 2651.
- [174] Adams, J.D., Jr.; Klaidman, L.K.; Odunze, I.N.; Shen, H.C.; Miller, C.A. *Mol. Chem. Neuropathol.*, **1991**, *14*, 213.
- [175] Jenner, P. *Lancet*, **1994**, *344*, 796.
- [176] Lohr, J.B.; Browning, J.A. *Psychopharmacol. Bull.*, **1995**, *31*, 159.
- [177] Liu, H.; Wang, H.; Shenvi, S.; Hagen, T.M.; Liu, R.M. *Ann. N. Y. Acad. Sci.*, **2004**, *1019*, 346.
- [178] Asensi, M.; Sastre, J.; Pallardo, F.V.; Lloret, A.; Lehner, M.; Garcia-de-la Asuncion, J.; Vina, J. *Methods Enzymol.*, **1999**, *299*, 267.
- [179] Moosmann, B.; Uhr, M.; Behl, C. *FEBS Lett.*, **1997**, *413*, 467.
- [180] Skaper, S.D.; Fabris, M.; Ferrari, V.; Dalle Carbonare, M.; Leon, A. *Free. Radic. Biol. Med.*, **1997**, *22*, 669.
- [181] Gutzmann, H.; Hadler, D. *J. Neural. Transm. Suppl.*, **1998**, *54*, 301.
- [182] Weyer, G.; Babej-Dolle, R.M.; Hadler, D.; Hofmann, S.; Herrmann, W.M. *Neuropsychobiology*, **1997**, *36*, 73.
- [183] Yamada, K.; Tanaka, T.; Han, D.; Senzaki, K.; Kameyama, T.; Nabeshima, T. *Eur. J. Neurosci.*, **1999**, *11*, 83.
- [184] Pluta, R. *Folia neuropathologica / Association of Polish Neuropathologists and Medical Research Centre, Polish Academy of Sciences*, **2000**, *38*, 188.
- [185] Thal, L.J.; Grundman, M.; Berg, J.; Ernstrom, K.; Margolin, R.; Pfeiffer, E.; Weiner, M.F.; Zamrini, E.; Thomas, R.G. *Neurology*, **2003**, *61*, 1498.
- [186] Gutzmann, H.; Kuhl, K.P.; Hadler, D.; Rapp, M.A. *Pharmacopsychiatry*, **2002**, *35*, 12.
- [187] Hampson, A.J.; Grimaldi, M.; Axelrod, J.; Wink, D. *Proc. Natl. Acad. Sci. USA*, **1998**, *95*, 8268.
- [188] Marsicano, G.; Wotjak, C.T.; Azad, S.C.; Bisogno, T.; Rammes, G.; Cascio, M.G.; Hermann, H.; Tang, J.; Hofmann, C.; Zieglgansberger, W.; Di Marzo, V.; Lutz, B. *Nature*, **2002**, *418*, 530.
- [189] Moosmann, B.; Skutella, T.; Beyer, K.; Behl, C. *Biol. Chem.*, **2001**, *382*, 1601.
- [190] Moosmann, B.; Behl, C. *Eur. J. Biochem.*, **2000**, *267*, 5687.
- [191] Munch, G.; Taneli, Y.; Schraven, E.; Schindler, U.; Schinzel, R.; Palm, D.; Riederer, P. *J. Neural Transm. Park Dis. Dement. Sect.*, **1994**, *8*, 193.
- [192] Shoda, H.; Miyata, S.; Liu, B.F.; Yamada, H.; Ohara, T.; Suzuki, K.; Oimomi, M.; Kasuga, M. *Endocrinology*, **1997**, *138*, 1886.
- [193] Stadtman, E.R. *Arch. Biochem. Biophys.*, **2004**, *423*, 2.
- [194] Moskovitz, J.; Berlett, B.S.; Poston, J.M.; Stadtman, E.R. *Proc. Natl. Acad. Sci. USA*, **1997**, *94*, 9585.
- [195] Moskovitz, J.; Bar-Noy, S.; Williams, W.M.; Requena, J.; Berlett, B.S.; Stadtman, E.R. *Proc. Natl. Acad. Sci. USA*, **2001**, *98*, 12920.
- [196] Moskovitz, J.; Flescher, E.; Berlett, B.S.; Azare, J.; Poston, J.M.; Stadtman, E.R. *Proc. Natl. Acad. Sci. USA*, **1998**, *95*, 14071.
- [197] Yermolaieva, O.; Xu, R.; Schinstock, C.; Brot, N.; Weissbach, H.; Heinemann, S.H.; Hoshi, T. *Proc. Natl. Acad. Sci. USA*, **2004**, *101*, 1159.
- [198] Ruan, H.; Tang, X.D.; Chen, M.L.; Joiner, M.L.; Sun, G.; Brot, N.; Weissbach, H.; Heinemann, S.H.; Iverson, L.; Wu, C.F.; Hoshi, T. *Proc. Natl. Acad. Sci. USA*, **2002**, *99*, 2748.
- [199] Grundman, M. *Am. J. Clin. Nutr.*, **2000**, *71*, 630S.

Copyright of *Mini Reviews in Medicinal Chemistry* is the property of Bentham Science Publishers Ltd. and its content may not be copied or emailed to multiple sites or posted to a listserv without the copyright holder's express written permission. However, users may print, download, or email articles for individual use.